Pitfalls and problems encountered in rat model sciatic nerve surgery

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ABSTRACT
When learning the basics of microsurgery, a trainee must be equipped with patience and perseverance in order to evolve. One must have the ground knowledge when it comes to peripheral nerve injury and nerve regeneration process in order to fully understand that the technique is vital for the outcome and final results. Furthermore, a trainee must practice on non-living tissue before performing successful in vivo operations and even in this case, one may be confronted with problems regarding the surgical technique. [1] The following article aims to reveal the main problems/mistakes when performing sciatic nerve surgery in an in vivo experimental model and the solutions for these problems.

INTRODUCTION
There are many experimental surgical projects performed on the sciatic nerve of the rat. [2] There are many reasons why researchers choose this model: high similitude in nerve regeneration between the rat and the human; the lab rat (most commonly used the Wistar rat) is more available compared to other species and easier to manipulate; the anatomy of the sciatic nerve of the rat offers easy access, without significant comorbidity; the diameter and the length of the nerve permits different types of experiments (direct suture, reconstruction of different gaps with nerve conducts). [3] The sciatic nerve has a length of ~2,5-3cm before it divides into 3 terminal branches - common peroneal nerve, tibial nerve and sural nerve.

Although rat’s sciatic nerve is one of the most used models for nerve regeneration it still may cause difficulties for an inexperienced surgeon. The problems one may encounter are related to anesthesia, [4] rat anatomy knowledge or microsurgical technique and each one of these issues may cause an enormous impact on the final outcome. Understanding the theoretical aspects of nerve regeneration [5],[6],[7] may help improve the surgical technique, especially when it comes to atraumatic manipulation.
Material and Method
For this review there were used Wistar rats in different microsurgery training laboratories. The rats varied in age and weight (between 250-350g or 7-9 weeks). The anesthesia protocol consisted of a mixture of ketamine (75mg/kg) and xylazine (10mg/kg) administered intraperitoneally. A precise scale was used to weigh every animal before administering the anesthesia. There were used proper microsurgical instruments as well as a microscope and appropriate sutures (8-0, 9-0, 10-0).

After shaving the dorsal aspect of the gluteal and thigh region, the rat is placed in prone position. An oblique-transverse skin incision is made and dissection along the biceps muscle fibers is performed in order to expose the underlying sciatic nerve. Mehedintu et al described different types of incisions to access the sciatic nerve. [7],[8] After nerve exposure, the rest of the operation is done under the microscope. This involves nerve dissection, nerve section and subsequent nerve repair. Proximal to its division, the sciatic nerve can be used for various operations (direct suture after section, autografting, using different nerve conduct for iatrogenic nerve defects).

Results
A frequent mistake of the in vivo experiment is the incorrect administration of the anesthesia, which must be very well dosed and not lead to overdose and rat death.

Aside from the anesthetic issues, the most frequent problems when repairing a nerve defect with autograft arise from lack of experience. [9] These are: identifying the sciatic nerve correctly, improper incision site, excessive nerve dissection for a better exposure, choosing an unsuitable location to create the nerve gap, as well as microsurgical technique errors (inappropriate cooptation, high suture tension). Inadequate sutures for the nerve size also influence the recovery results.
Distal incision beyond the division of the sciatic nerve (marked with arrow)

Axoplasmatic protrusion (arrow) in the proximal nerve suture due to insufficient proximal dissection

Identifying the sciatic nerve correctly poses no problem if the skin incision site and the muscle dissection is performed in the right area and in a proper plane. A distal incision (too lateral and caudal), however, may create confusion in case not all branches are visible, thus mistaking the sciatic nerve to one of its branches. A contributing factor to this problem is represented by the placing of the rat in a slight prone-lateral position (instead of the custom prone position). These branches are half or even 1/3 of the sciatric nerve diameter (~1mm). To prevent this situation, the skin incision should be placed more cranial and medial, parallel and 1cm inferior to the femur, with the rat in prone position. When suspecting that the first incision is inappropriate, with exposure of just one branch of the sciatic nerve, extending the incision more proximal can solve the problem by identifying the correct nerve.

Another technical error is creating the nerve gap
too proximal (or not dissecting enough of the sciatic nerve from the muscle tissue in the proximal part). After nerve sectioning, the elastic fibers in the structure of the nerve determine retraction of the proximal stump in a plane which is hard to access, thus making the nerve suture difficult to perform. In order to prevent this problem, a proper dissection and the selection of the central area of the sciatic nerve to create the gap is recommended. The consequences of suturing a retracted stump are: inappropriate traction of the nerve (which risks injuring the nerve fascicles), unmatched fascicles in the suture (the surgeon is no longer able to observe the epineural vessels to guide a correct cooptation), axoplasmatic protrusion due to excessive traction and uneven suture. In all cases, inappropriate dissection results in prolonged operating times.

Creating the nerve gap too distal (close to the division of the sciatic nerve) poses a problem for nerve suturing because an epineural suture tends to shred the common sheath of the branches, dividing them into 3 separate nerves (which require individual repair). The ideal distal sectioning site is ~0.7cm proximal to the visible division of the sciatic nerve in order to have enough connective tissue for a strong nerve suture.

**Discussion**

It is recommended not to operate on rats lighter than 250g and younger than 7 weeks of age. Otherwise the nerve diameter might be smaller than 2mm, requiring adequate microsurgery equipment as well as a greater experience in nerve surgery. Above 350g or 9 weeks of age, the animals require more substance to induce anesthesia, without any extra benefits.

Multiple administration of anesthesia might lead to overdose, causing respiratory distress and in the end exitus. For this reason, if the anesthesia doesn’t work accordingly from the beginning with the calculated dose, the operation should be postponed and the anesthetic options should be revised.

The iatrogenic nerve gap should be placed centrally, after exposing the sciatic nerve from its origin to its division.

**Conclusions**

Microsurgery requires adequate instruments and special training. Calculating the exact quantity of anesthetic reduces the death risk. Experience and attention are key factors in avoiding different complications related to nerve surgery. The microsurgical technique involves an atraumatic manipulation of the tissue, correct cooptation without suture tension and right alignment of the nerve fascicles. Respecting these rules results in a good nerve regeneration, with positive clinical outcomes.

**References**


